



Written By: **Sam Savage** Published Date: **May 16, 2008** Last Edited: **May 16, 2008**

By Hu, Dan-Ning Simon, John D; Sarna, Tadeusz

ABSTRACT The mammalian eye consists of several layers of pigmented tissues that contain melanin. The eye is a unique organ for pigment cell research because one can isolate and compare melanosomes from different tissues and embryonic origins. Retinal, iris and ciliary pigment epithelial cells are derived from the neural ectoderm, more specifically from the extremity of the embryonic optical cup, which is also the origin of the retina. In contrast, the pigment-generating cells in the choroid and in the stroma of the iris and ciliary body, uveal melanocytes, are developed from the neural crest, the same origin as the melanocytes in skin and hair. This review examines the potential functions of ocular melanin in the human eye. Following a discussion of the role of melanins in the pigment epithelium and uveal melanocytes, three specific topics are explored in detail-photo-screening protective effects, biophysical and biochemical protective effects, and the biologic and photobiologic effects of the two main classes of melanins (generally found as mixtures in ocular melanosomes)- eumelanin and pheomelanin.

INTRODUCTION

The wall of the human eye consists of three layers, the transparent cornea and opaque white sciera, the uveal tract and the retina (1). The uveal tract, a highly vascularized connective tissue, is further composed of three parts, from anterior to posterior-the iris, the ciliary body and the choroid. The choroid supports and nourishes the retina, which is located on the inner side of the choroid. The retina further consists of two layers-the retinal pigment epithelium (RPE) and the neural retina. The neural retina contains photoreceptor cells, which are involved in the primary processes of visual transduction, and other neurons, which encode and transfer the visual information to the brain. The RPE (derived from the neuroectoderm), a monolayer of postmitotic pigment cells that lies between the uveal tract and the neural retina, is

 \oslash

responsible for important metabolic support for the entire retina and is involved in phagocytosis of the photoreceptor outer segment disks, which are constantly being shed (2). The RPE extends to and is contiguous with the iris pigment epithelium (IPE) and ciliary pigment epithelium. A sagittal horizontal section of the adult human eye is shown in Fig. 1.

Melanin is found in several of these tissues. Pigmented cells are of two different types-the uveal melanocytes located in the uveal tract, and the pigment epithelial cells (1-4). The uveal melanocytes in the uveal tract are derived from the neural crest and can be divided into iridal, ciliary and choroidal melanocytes (1-4). Melanocytes in the iris and ciliary body are located in the stroma. Melanin is also found in all three of the pigment epithelium cell types, of which the RPE is the most studied.

The function of melanin in these various tissues is not fully elucidated. Melanin tends to protect the eye against several ocular diseases that can cause blindness, including uveal melanoma and age- related macular degeneration (AMD) (4-6). However, the exact mechanism by which melanin protects the eye, whether the protective function depends on the type of the melanin, and whether the melanin- related protection changes with age, remains mostly unknown. This article examines the current hypotheses for the role melanin plays in the physiology and pathology of the eye. Because many of these hypothesized roles are linked to its interaction with light, we first summarize the accessibility and exposure of ocular different pigment cells to sunlight and UV radiation (7).

Environmental light impinging on the eye consists of the visible and UV regions of the electromagnetic spectrum. The UV region is further subdivided into UVA, UVB and UVC. According to the International Commission on Illumination, the wavelength ranges of the regions in the UV are-UVC: 100-280 nm, UVB: 280-315 nm and UVA: 315-400 nm. Definitions based on biologic effects modify these rangesUVC: 180-290 nm, UVB: 290-320 nm and UVA: 320400 nm. UVC in sunlight is normally completely screened by stratospheric ozone, but it is important to note that artificial light sources can also produce UVC.

Not all wavelengths of light impinging on the surface of the eye illuminate the various melanin-containing cells in the eye. The iridal melanocytes are located behind the cornea and anterior chamber (containing the aqueous humor). The cornea is transparent to visible light, but it absorbs all of the UVC, part of the UVB (22- 73% at 320-300 nm) and a very small amount of UVA (6-20% at 400-330 nm) (8). Therefore, in vivo the iridal melanocytes are exposed only to visible light, UVA and some of the UVB spectrum. The ciliary body and choroidal melanocytes are covered internally by the retina and densely pigmented ciliary and retinal pigment epithelia and externally by thick and nontransparent sciera. In infancy and in early childhood, there is a window of transmission of nearly 8% of UV radiation around 320 nm through the lens, and about 30% of the transmitted UV is absorbed by the RPE before impinging upon the uveal melanocytes (8,9). As a result of the transmission properties of the cornea and lens, only visible light reaches the RPE in the adult human eye (7).

Figure 1. A sagittal horizontal section of the adult human eye. Reprinted with permission from http://www.webvision.med.utah.edu.

The remainder of this review is organized as follows. First we briefly review the chemistry of melanins and the melanogenesis of ocular melanosomes. Second, we focus on the iris, examining the relationship between iris color and melanin composition, and eye diseases. This is followed by a general discussion of the role ocular melanin might play in the physiology and pathology of the eye. Three specific topics are explored-photo-screening protective effects, biophysical and biochemical protective effects, and the biologic and photobiologic effects of the two main classes of melanins (generally found as mixtures in ocular melanosomes)- eumelanin and pheomelanin.

MELANIN AND OCULAR MELANOGENESIS

There are different types of melanin present in the pigment epithelia and uveal melanocytes. The pigment epithelium is densely pigmented in all races and in all eye colors. Melanin in the pigment epithelium is mainly eumelanin, which is a brownblack substance derived from tyrosine or dopa. Eumelanin is formed in a series of oxidation and tautomerization reactions catalyzed by several enzymes, with the end product being a complex oligomeric material exhibiting a distinct particle nature (10-13). Key intermediates in the biosynthesis of eumelanin are 5,6-dihydroxyindole and 5,6- dihydroxyindole-2-carboxylic acid, as well as their oxidized forms.

Formation of melanosomes occurs in the RPE early in fetal development, then ceases within a few weeks (14). Polymerization of melanin within these melanosomes continues until, at approximately 2 years of age in humans, the RPE contains only mature melanosomes (14). Whether melanogenesis occurs in the RPE after approximately 2 years has not been definitely established. Premelanosomes, or partially melanized melanosomes, which are indicative of ongoing melanogenesis, have not been observed in adult human RPE. In addition, very little or no tyrosinase activity could be detected in adult bovine RPE cells (15,16). The melanin content of the RPE decreases significantly in aged human eyes (17-20). Therefore, melanin biosynthesis either is absent in adult human RPE cells or occurs only at a very slow rate; and whether there is turnover of RPE melanosomes remains unknown.

In uveal melanocytes, the quality and quantity of melanin vary with race and iris color. In the uveal pigments, pheomelanin is often present in addition to eumelanin (21-23). Pheomelanin is a lighter colored, yellowish pigment that is formed when cysteine or glutathione is present during the oxidation stage of dopa (24). 1,4- benzothiazynylalanine, derived from cysteinyldopas, is proposed to be a key intermediate in the biosynthesis of pheomelanin (24). The quantity of uveal melanin in eyes with dark-colored irides is greater than that in light-colored eyes (21,23,25). Uveal melanocytes contain both eumelanin and pheomelanin. In cells from eyes with dark-colored irides (brown and dark brown in color), the amount of eumelanin and the ratio of eumelanin/pheomelanin is significantly greater than that from eyes with light-colored irides (hazel, green, yellow-brown and blue in color) (23). The quantity of pheomelanin in uveal

melanocytes from eyes with light-colored irides is slightly greater than that from dark-colored irides, although the difference is not statistically significant (23).

The ocular melanin content differs among species. For example, Liu et al. (22) reported that pheomelanin content in bovine eyes is low in the choroid and RPE and moderate in the iris (containing both iridal melanocytes and IPE). In cultured human uveal melanocytes, the quantity and type of melanin in iridal melanocytes are not significantly different from that in choroidal melanocytes (23).

Both uveal melanocytes and pigment epithelium cells can be isolated and cultured in vitro (26-30). Human uveal melanocytes produce melanin to maintain a constant level of melanin in vitro. Cultured uveal melanocytes isolated from eyes with different iris colors maintain their inherent capacity for melanogenesis (31). Adult human pigment epithelium cells do not produce melanin in vitro and perhaps not in vivo either (32). The melanin content of cultured RPE decreases rapidly and in proportion to cell division. No melanin production could be demonstrated in cultured RPE under standard culture circumstances (27,30,32-34). Several authors have reported that cultured human adult RPE may produce melanin under special circumstances or when induced by certain stimulators (35-37). These reports have not provided a quantitative measurement of melanin in the cultured RPE cells, have proven difficult to replicate by others, and have not established that the pigment produced is the same as that naturally found in the cells. IRIS-RELATIONSHIPS BETWEEN COLOR, MELANIN COMPOSITION AND DISEASE

The IPE is located at the posterior surface of the iris. The IPE is pigmented in all races and colors. The pigment in the IPE provides only a background tint, receiving and reflecting light only through the filter of stroma arranged in front of this tissue (21,38- 41). The iris color is determined by the variation in pigmentation of the melanocytes in the stroma.

The quantity and types of melanin in the iridal melanocytes vary with iris color (21,23,42). However, it is important to emphasize that the iris color visible through the cornea results from different optical phenomena, such as multiple light scattering on pigment granules and other components of the connective tissue forming the stroma, as well as light absorption by various chromophores (26). Studies of human donor eyes under light and electron microscopes revealed that the difference in iris color is determined by the variation of the melanosome structure and composition within the iridal melanocytes, not by the number of iridal melanocytes present (38,40,41). Darker indes have larger melanin granules and greater granule density (38). In pathologic conditions, e.g. albinism, where the melanin content in the pigment epithelium is markedly decreased or even absent, very light-colored irides may vary from yellow to a pink color.

The incidence of two important eye diseases, uveal melanoma and AMD, appears to be correlated with the color of the iris. Uveal melanoma is the most common intraocular malignant tumor in human adults. A population-based study on the relationship between racial/ ethnic group and incidence of uveal melanoma found that the incidence of uveal melanoma is highest in non-Hispanic whites, followed by Hispanics,

Asians and blacks, with a white/black incidence ratio of uveal melanoma of 18:1 (43). These epidemiologic data suggest that the light-colored eye is at higher risk for the occurrence of uveal melanoma. In fact, several studies have shown that light-colored irides (blue, hazel, etc.) have a higher incidence of uveal melanoma (44-46). Recently, a meta-analysis based on 10 studies (1732 cases) revealed that a blue or gray iris is a statistically significant risk factor for the development of uveal melanoma (47).

AMD is a common ocular disease that is the major cause of blindness among the elderly in developed countries. AMD is at least an order of magnitude (48) more prevalent in the white population than in darkly pigmented races, suggesting that melanin may be protective against AMD development (49-52). Several authors have found an association between light-colored irides and the occurrence or progress of AMD, although the relationship between iris color and AMD is not so conclusive as that in uveal melanoma (20,50,53-57).

PROTECTIVE EFFECTS OF OCULAR MELANIN

The detrimental effects of UV radiation are a cause of the cellular gene mutation that leads to cutaneous melanoma. Reactive oxygen species (ROS), both UV-induced and biochemically produced, also play a role in the malignant transformation of uveal melanocytes. ROS can be either stable diamagnetic molecules or free radicals; when they are produced in the choroid and RPE they can damage the RPE and lead to the degeneration of photoreceptors in the neural retina, e.g. AMD.

The protective effects of melanin on the ocular cells and tissues occur by both physical and biochemical mechanisms; the pigment acts as a photo-screen and as an antioxidant, respectively (6). The photoscreening effect, purely physical in nature, dominates in the anterior segment (the iris), which is exposed to sunlight and UV radiation. The posterior segment is exposed to limited amounts of light and UV radiation. Visible light reaches RPE melanosomes but the exposure of choroidal melanosomes to light is very limited. In these regions the sole mechanism of protection must be biochemical (58). We now examine each of these effects in more detail.

Photo-screening protective effects

Melanin absorbs near-infrared, visible light and UV radiation with absorption increasing at the shorter wavelengths (6). In the anterior segment of the eye, the pigment epithelium and the melanocytes in the iris absorb and block both visible light and UV radiation, thus protecting the rest of the eye from the deleterious effects of these wavelengths. A significant amount of light escapes the absorption by photoreceptor cells, and so even in the posterior segment of the eye (e.g. RPE), melanosomes absorb light. In fact, absorption by the RPE is believed to aid in minimizing spurious signals that may appear because of light reflection and scatter from the fundus (5). Based on experimental measurements, it has been estimated that the absorbance of the RPE, resulting mostly from absorption by RPE melanosomes, is in the range 0.2-0.9. Thus, the amount of light reaching the choroidal melanocytes is much lower than that

reaching the iris and RPE, but remains a concern. These uveal melanosomes may still act as a photo-screen, but this may not be the major role they play here in mitigating the onset or progression of uveal melanoma or AMD.

Paradoxically, it has been reported that solar radiation causes a decrease in the incidence of uveal melanoma (59). This is consistent with the dual effect of UV radiation on the occurrence of other malignant tumors. Recently, it has been reported that solar radiation reduces the risk and/or mortality of various systemic malignant tumors that are not exposed to sunlight, e.g. non- Hodgkin's lymphoma, and prostate, breast, colon and ovarian cancers. These beneficial effects occur because UV radiation increases vitamin D synthesis in the skin; vitamin D then converts to 1,25- dehydroxyvitamin D3, which inhibits growth and induces apoptosis in various malignant tumor cells both in vitro and in experimental animal models. Therefore, sunlight has dual effects on malignant tumors-a direct mutagenic effect on tissues exposed to the sunlight and an indirect protective effect on tissues not exposed to sunlight (59).

Cutaneous and conjunctival melanocytes are mainly exposed to solar radiation, and in their tissues the direct effect of UV radiation predominates and causes an increase in tumor incidence with decreasing latitude (increasing solar radiation). Uveal melanocytes, mainly the choroidal and ciliary body melanocytes, are not directly exposed to solar radiation, so no direct effect of solar radiation would be expected to occur in these locations. Therefore, the indirect protective effect of solar radiation causes a decrease in uveal melanoma (59).

The lower incidence of AMD in darkly pigmented eyes may be related to lower light intensity that is transmitted to the retina. This is because darkly pigmented indes (with more iridal melanin) will more efficiently attenuate the light that reaches the eye fundus. The spectrum of light transmitted by differently pigmented indes depends on the color of the indes. So if one hypothesizes that the actual damage that triggers the cellular processes leading to AMD is in the RPE, then melanin in the RPE can offer some protection against light-related phenomena. Indeed, there is a growing body of experimental evidence suggesting that AMD actually originates in the pathologic changes in the RPE (60).

The photo-screening effect of melanin can also play a role in melanoma of the iris. Iris melanoma is much rarer and less malignant than ciliary body and choroidal melanomas. The melanocytes of the iris are located in the eye's anterior surface and exposed to solar radiation. Iris melanoma tends to occur in the inferior sector of the iris, where exposure to sunlight is the greatest (61), indicating that its occurrence is related to exposure to UV radiation. The lower incidence of iridal melanoma in dark-colored eyes (61) might be related to the photo-screening effect provided by their more abundant iridal melanin.

Biophysical/biochemical protective effects

The choroid, located in the posterior segment of the eye, is highly vascularized and therefore is at elevated risk of experiencing significant oxidative stress. Choroidal melanin, an antioxidant and a weak free radical

scavenger, may deactivate ROS and protect the retina from oxidative damage (30,59). However, with age, the constant exposure of pigment cells to high levels of oxygen may diminish the antioxidant properties of melanin. In this case, melanin may even become a pro-oxidant, which may lead to the damage of photoreceptors and cause AMD (5,30,59). Uveal melanocytes in eyes with dark-colored irides contain a greater amount of melanin and therefore can resist ROS and protect the tissues until a later point in the aging process. This effect could explain the decrease in the incidence of AMD in the dark-colored eye.

Biochemical protective effects in the RPE may also play a role in the occurrence of AMD. Melanin in the RPE can act against ROS and protect the neural retina (62,63). With age, the constant exposure of the RPE to high levels of oxygen and light might diminish the antioxidant properties of melanin (64-67). Under these conditions melanin may become pro-oxidant, adding to the accumulation of the singlet-oxygen-producing pigment lipofuscin in the cytoplasm of aged RPE cells and ultimately leading to AMD (5,6,14,17-19,30,62,63,68). Uveal melanin, especially in the ciliary body and choroid, can also protect melanocytes from oxidative stress and reduce the malignant transformation of uveal melanocytes. Melanocytes in dark-colored eyes have a high quantity of melanin, which is more protective than that in light-colored eyes, consistent with the higher incidence of uveal melanoma in the light-colored eye (23,47,59).

BIOLOGIC AND PHOTOBIOLOGIC EFFECTS OF EUMELANIN COMPARED TO PHEOMELANIN

Several studies have compared the reactivity of eumelanin and pheomelanin and found that both melanins act as free radical scavengers and inhibit UV-induced lipid peroxidation (69-72). However, the antioxidant properties of melanin are related to the type of melanin-the greater the ratio of eumelanin to pheomelanin, the more antioxidative the pigment (69,70). Pheomelanin complexed with Fe (III) stimulates UV-induced lipid peroxidation, whereas eumelanin does not (71,72). Cultured melanocytes with high levels of eumelanin show a better survival rate after irradiation with UVB (73). UV irradiation of melanin also generates ROS, and this photosensitization is greater for pheomelanin than for eumelanin (72,73).

Takeuchi et al. (74) examined the induction of DNA lesions and apoptosis upon UV exposure of congenic mice with black, yellow and albino coats. UVB-induced cyclobutane dimerization and apoptosis measured by sunburn cells or keratinocytes containing active caspase- 3 was strain independent. Combining the results of measurements on TUNEL-positive cells with the concentration of pigments in different mice revealed that compared to eumelanin, the presence of pheomelanin induces a three-fold greater activity. This result strongly supports the conclusion that pheomelanin sensitizes apoptosis (via caspase-3 activation) in adjacent cells at a frequency greater than that induced by direct DNA absorption. Studies using free-electron laser photoelectron emission microscopy, femtosecond time-resolved absorption spectroscopy and electron spin resonance oximetry reveal that unlike eumelanosomes, pheomelanosomes exhibit a second threshold potential of 3.8 eV, corresponding to photons with wavelengths as long as 326 nm (75,76). The data suggest that pheomelanosomes may be more susceptible to adverse reactions induced by solar radiation.

Uveal melanin in dark-colored eyes contains more eumelanin than that in light-colored eyes (23). Because both melanins are protective and eumelanin is less photoreactive than pheomelanin, the high level of eumelanin in dark-colored eyes suggests that dark- colored eyes would have a lower incidence of uveal melanoma and AMD, consistent with the results of epidemiologic studies (43,47-52).

REFERENCES

1. Snell, R. S. and M. A. Lemp (1998) Clinical Anatomy of the Eye. Wiley, New York.

2. Bumstead, K. M. and C. J. Barnstable (2000) Dorsal retinal pigment epithelium differentiate as neural retina in microphthalmia(mi/mi) mouse. Invest. Ophthalmol. Vis. Sci. 41, 903- 908.

3. Dulac, C. (1993) The embryonic development of melanocytes and its pathology. M. S-Med Sci. 9, 417-424.

4. Hu, D. N. (2005) Photobiology of ocular melanocytes and melanoma. Photochem. Photobiol. 81, 506-509.

5. Sarna, T. (1992) Properties and function of the ocular melanin: A photobiophysical view. J. Photochem. Photobiol. 12, 215- 258.

6. Sarna, T. and H. A. Swartz (1998) The physical properties of melanin. In The Pigment System: Physiology and Pathophysiology (Edited by J. J. Nordland, R. E. Boissy, V. J. Hearing, R. A. King and J.-P. Ortonne) pp. 333-358. Oxford University Press, Oxford.

7. Sliney, D. H. (2005) Exposure geometry and spectral environment determine photobiological effects on the human eye. Photochem. Photobiol. 81, 483-489.

8. Kurzel, R. B., M. L. Wolbarsht and B. S. Yamanashi (1977) Ultraviolet radiation effects on the human eye. Photochem. Photobiol. Rev. 2. 133-167.

9. Singh, A. D., I. G. Rennie, S. Seregard, M. Giblin and J. McKenzie (2004) Sunlight exposure and pathogenesis of uveal melanoma. Surv. Ophthalmol. 49, 419-428.

10. Wakamatsu, K. and S. Ito (2002) Advanced chemical methods in melanin determination. Pigment Cell Res. 15, 174-183.

11. Clancy, C. M. R. and J. D. Simon (2001) Ultrastructural organization of eumelanin from Sepia officinalis measured by atomic force microscopy. Biochemistry 40, 13353-13360.

12. Liu, Y. and J. D. Simon (2003) Isolation and biophysical studies of natural eumelanins: Applications of imaging technologies and ultrafast spectroscopy. Pigment Cell Res. 16, 606-618.

13. Meredith, P. and T. Sarna (2006) The physical and chemical properties of eumelanin. Pigment Cell Res. 19, 572-594.

14. Boulton, M. (1998) Melanin and the retinal pigment epithelium. In The Pigment Epithelium. Function and Disease (Edited by M. F. Marmor and T. Wolfensberger), pp. 68-85. Oxford University Press, New York.

15. Dryja, T. P., M. O'Neil-Dryja, J. M. Pawelek and D. M. Albert (1978) Demonstration of tyrosinase in the adult bovine uveal tract and retinal pigment epithelium. Invest. Ophthalmol. Vis. Sci. 17, 511-514.

16. Nakazawa, M., M. Tsuchiya, S. Hayasaka and K. Mizuno (1985) Tyrosinase activity in the uveal tissue of the adult bovine eye. Exp. Eye Res. 41, 249-258.

17. Sarna, T., J. M. Burke, W. Korytowski, M. Rozanowska, C. M. Skumatz, A. Zareba and M. Zareba (2003) Loss of melanin from human RPE with aging: Possible role of melanin photooxidation. Exp. Eye Res. 76, 89-98.

18. Boulton, M. and P. Dayhaw-Barker (2001) The role of the retinal pigment epithelium: Topographical variation and ageing changes. Eye 15, 384-389.

19. Schmidt, S. Y. and R. D. Peisch (1986) Melanin concentration in normal human retinal pigment epithelium: Regional variation and age-related reduction. Invest. Ophthalmol. Vis. Sci. 27, 10631067.

20. Weiter, J. J., F. C. Delori, G. L. Wing and K. A. Fitch (1985) Relationship of senile macular degeneration to ocular pigmentation. Am. J. Ophthalmol. 99, 185-187.

21. Prota, G., D.-N. Hu, M. R. Vincensi, S. A. McCormick and A. Napolitano (1998) Characterization of melanins in human irides and cultured uveal melanocytes from eyes of different colors. Exp. Eye Res. 67, 293-299.

22. Liu, Y., L. Hong, K. Wakamatsu, S. Ito, B. B. Adhyaru, C. Y. Cheng, C. R. Bowers and J. D. Simon (2005) Comparisons of the structural and chemical properties of melanosomes isolated from retinal pigment epithelium, iris and choroid of newborn and mature bovine eyes. Photochem. Photobiol. 81, 510-516.

23. Wakamatsu, K., D. N. Hu, S. A. McCormick and S. Ito (2008) Characterization of melanin in human iridal and choroidal melanocytes from eyes with various colored irides. Pigment Cell Res. 21, 97-105.

24. Slominski, A., D. J. Tobin, S. Shibahara and J. Wortsman (2004) Melanin pigmentation in mammalian skin and its hormonal regulation. Physiol. Rev. 84, 1155-1228.

25. Wielgus, A. R. and T. Sarna (2005) Melanin in human irides of different color and age of donors. Pigment Cell Res. 18, 454-464.

26. Hu, D. N., R. Ritch, S. A. McCormick and K. Pelton-Henrion (1992) Isolation and cultivation of human iris pigment epithelium. Invest. Ophthalmol. Vis. Sci. 33, 2443-2453.

27. Flood, M. T., P. Gouras and H. Kjeldbye (1980) Growth characteristics and ultrastructure of human retinal pigment epithelium in vitro. Invest. Ophthalmol. Vis. Sci. 19, 1309-1320.

28. Hu, D. N., S. A. McCormick and R, Ritch (1993a) Studies of human uveal melanocytes in vitro: Growth regulation of cultured human uveal melanocytes. Invest. Ophthalmol. Vis. Sci. 34, 22202227.

29. Hu, D. N. (2000) Regulation of growth and melanogenesis of uveal melanocytes. Pigment Cell Res. 13 (Suppl. 8), 81-86.

30. Hu, D. N., H. Savage and J. E. Roberts (2002) Uveal melanocytes, ocular pigment epithelium and Mueller cells in culture: In vitro toxicology. Int. J. Toxicol. 21, 465-472.

31. Hu, D. N., S. A. McCormick, S. J. Orlow, S. Rosemblat, A. Y. Lin and K. Wo (1995) Melanogenesis in cultured human uveal raelanocytes. Invest. Ophthalmol. Vis. Sci. 36, 931-938.

32. Lu, F., D. Yan, X. Zhou, D. N. Hu and J. Qu (2007) Expression of melanin-related genes in cultured adult human retinal pigment epithelium and uveal melanoma cells. Mol. Vis., 13, 2066-2072.

33. Newsome, D. A. (1983) Retinal pigment epithelium culture. Current application. Trans. Ophthalmol. Soc. UK 103, 458-466.

34. Smith-Thomas, L., P. Richardson, A. J. Thody, A. Graham, I. Palmer, L. Flemming, M. A. Parsons, I. G. Rennie and S. MacNeil (1996) Human ocular raelanocytes and retinal pigment epithelial cells differ in their melanogenic properties in vivo and in vitro. Curr. Eye Res. 15, 1079-1091.

35. Kurtz, M. J. and R. B. Edwards (1991) Influence of bicarbonate and insulin on pigment synthesis by cultured adult human retinal pigment epithelial cells. Exp. Eye Res. 53, 681-684.

36. Pfeffer, B. A. (1991) Improved methodology for cell culture of human and monkey retinal pigment epithelium. In Progress in Retinal Research (Edited by N. Osborne and G. Chader), pp. 251291. Pergamon Press, Oxford.

37. Rak, D. J., K. M. Hardy, G. J. Jaffe and B. S. McKay (2006) Ca(+ +)-switch induction of RPE differentiation. Exp. Eye Res. 82, 648-656.

7/3/2019

38. Eagle, R. C., Jr (1988) Iris pigmentation and pigraented lesions: An ultrastructural study. Trans. Am. Ophthalmol. Soc. 84, 581687.

39. Feeney, L., J. A. Grieshaber and M. J. Hogan (1965) Studies on human ocular pigment. In Eye Structure. II. Symposium (Edited by J. W. Rohen), pp. 535-548. Schattauer-Verlag, Stuttgart.

40. Imesch, P. D., C. D. Bindley, Z. Khademian, B. Ladd, R. Gangnon, D. M. Albert and I. H. L. Wallow (1996)
Melanocytes and iris color: Electron microscope findings. Arch. Ophthalmol, 114, 443- 447. 41. Wilkerson, C.
L., N. A. Syed, M. R. Fisher, N. L. Robinson, I. H. L. Wallow and D. M. Albert (1996) Melanocytes and iris color:
Light microscopic findings. Arch. Ophthalmol. 114, 437-442.

42. Wakamatsu, K., R. Kavanagh, A. L. Kadekaro, S. Terzieva, R. Sturm, S. Leachman, A. Abdel-Malek and S. Ito (2006) Diversity of pigmentation in cultured human melanocytes is due to differences in the type as well as quantity of melanin. Pigment Cell Res. 19, 154- 162.

43. Hu, D. N., G. P. Yu, S. A. McCormick, S. Schneider and P. T. Finger (2005) Population-based incidence of uveal melanoma in various races and ethnic groups. Am. J. Ophthalmol. 140, 612617.

44. Holly, E. A., D. A. Aston, D. H. Char, J. J. Kristiansen and D. K. Ahn (1990) Uveal melanoma in relation to ultraviolet light exposure and host factors. Cancer Res. 50, 5773-5777.

45. Pane, A. R. and L. W. Hirst (2000) Ultraviolet light exposure as a risk factor for ocular melanoma in Queensland, Australia. Ophthalmic Epidemiol. 7, 159-167.

46. Vajdic, C. M., A. Kricker, M. Giblin, J. McKenzie, J. Aitken, G. G. Giles and B. K. Armstrong (2001) Eye color and cutaneous nevi predict risk of ocular melanoma in Australia. Int. J. Cancer 92, 906- 912.

47. Weis, E., C. P. Shah, M. Lajous, J. A. Shields and C. L. Shields (2006) The association between host susceptibility factors and uveal melanoma: A meta-analysis. Arch. Ophthalmol. 124, 54-60.

48. Age-Related Eye Disease Study Research Group (2000) Risk factors associated with age-related macular degeneration: A case- control study in the age-related eye disease study: Age-Related Eye Disease Study Report Number 3. Ophthalmology 107, 22242232.

49. Friedman, D. S., J. Katz, N. M. Bressler, B. Rahmani and J. M. Tielsch (1999) Racial differences in the prevalence of age- related macular degeneration: The Baltimore Eye Survey. Ophthalmology 106, 1049-1055.

50. Klein, R., M. L. Rowland and M. I. Harris (1995) Racial/ ethnic differences in age-related maculopathy. Third National Health and Nutrition Examination Survey. Ophthalmology 102, 371-381.

7/3/2019

Role of Ocular Melanin in Ophthalmic Physiology and Pathology - Redorbit

51. Klein, R., B. E. Klein, E. K. Marino, L. H. Kuller, C. Furberg, G. L. Burke and L. D. Hubbard (2003) Early agerelated maculopathy in the Cardiovascular Health Study. Ophthalmology 110, 25-33.

52. Klein, R., B. E. Klein, M. D. Knudtson, T. Y. Wong, M. F. Cotch, K. Liu, G. Burke, M. F. Saad and D. R. Jacobs Jr (2006) Prevalence of age-related macular degeneration in 4 racial/ethnic groups in the multi-ethnic study of atherosclerosis. Ophthalmology 113, 373-380.

53. Frank, R. N., J. E. Puklin, C. Stock and L. A. Canter (2000) Race, iris color, and age-related macular degeneration. Trans. Am. Ophthalmol. Soc. 98, 109-115.

54. Hyman, L. G., A. M. Lilienfeld, F. L. Ferris 3rd and S. L. Fine (1983) Senile macular degeneration: A casecontrol study. Am. J. Epidemiol. 118, 213-227.

55. Mitchell, P., W. Smith and J. J. Wang (1998) Iris color, skin sun sensitivity, and age-related maculopathy: The Blue Mountains Eye Study. Ophthalmology 105, 1359-1363.

56. Mitchell, P., J. J. Wang, S. Foran and W. Smith (2002) Five- year incidence of age-related maculopathy lesions: The Blue Mountains Eye Study. Ophthalmology 109, 1092-1097.

57. Sandberg, M. A., A. R. Gaudio, S. Miller and A. Weiner (1994) Iris pigmentation and extent of disease in patients with neovascular age-related macular degeneration. Invest. Ophthalmol. Vis. Sci. 35, 2734-2740.

58. Hong, L., J. D. Simon and T. Sarna (2006) Melanin structure and the potential functions of uveal melanosomes. Pigment Cell Res. 19, 465-466.

59. Hu, D. N., S. A. McCormick and G. P. Yu (2008) Latitude and incidence of uveal melanoma. Ophthalmology in press.

60. Bok, D. (2005) Evidence for an inflammatory process in age- related macular degeneration gains new support. Proc. Natl Acad. Sci. USA 102, 7053-7054.

61. Shields, J. A. and S. C. Shields (1992) Intraocular Tumor: A Text and Atlas, pp. 54-306. W. B. Saunders, Philadelphia.

62. Peters, S., T. Lamah, D. Kokkinou, K. U. Bartz-Schmidt and U. Schraermeyer (2006) Melanin protects choroidal blood vessels against light toxicity. Z. Naturforsch. 61, 427-433.

63. Wang, Z., J. Dillon and E. R. Gaillard (2006) Antioxidant properties of melanin in retinal pigment epithelial cells. Photochem. Photobiol. 82, 474-479.

64. Zareba, M., M. W. Raciti, M. M. Henry, T. Sarna and J. M. Burke (2006) Oxidative stress in ARPE-19 cultures: Do melanosomes confer cytoprotection? Free Radic. Biol. Med. 40, 87-100.

65. Zareba, M., T. Sarna, G. Szewczyk and J. M. Burke (2007) Photobleaching of melanosomes from retinal pigment epithelium: II. Effects on the response of living cells to photic stress. Photochem. Photobiol. 83, 925-930.

66. Zadlo, A., M. Rozanowska, J. M. Burke and T. Sarna (2007) Photobleaching of retinal pigment epithelium melanosomes reduces their ability to inhibit iron-induced peroxidation of lipids. Pigment Cell Res. 20, 52-60.

67. Burke, J. M., M. M. Henry, M. Zareba and T. I. Sarna (2007) Photobleaching of melanosomes from retinal pigment epithelium: I. Effects on protein oxidation. Photochem. Photobiol. 83, 920-924.

68. Weiter, J. J., F. C. Delori, G. L. Wing and K. A. Fitch (1986) Retinal pigment epithelial lipofuscin and melanin and choroidal melanin in human eyes. Invest. Ophthalmol. Vis. Sci. 27, 145-152.

69. Chedekel, M. R., P. P. Agin and R. M. Sayre (1980) Photochemistry of pheomelanin: Action spectrum for superoxide production. Photochem. Photobiol. 31, 553-555.

70. Chedekel, M. R., S. K. Smith, P. W. Post, A. Pokora and D. L. Vessell (1978) Photodestruction of pheomelanin: Role of oxygen. Proc. Natl Acad. Sci. USA 75, 5395-5399.

71. Krol, E. S. and D. C. Liebler (1998) Photoprotective actions of natural and synthetic melanins. Chem. Res. Toxicol. 11, 1434- 1440.

72. Takeuchi, S., W. Zhang, K. Wakamatsu, S. Ito, V. J. Hearing, K. H. Kraemer and D. E. Brash (2004) Melanin acts as a potent UVB photosensitizer to cause a novel mode of cell death in murine skin. Proc. Natl Acad. Sci. USA 101, 15076-15081.

73. de Leeuw, S. M., N. P. Smit, M. Van Veldhoven, E. M. Pennings, S. Pavel, J. W. Simons and A. A. Schothorst (2001) Melanin content of cultured human melanocytes and UV-induced cytctoxicity. J. Photochem. Photobiol. B, Biol. 61, 106-113.

74. Takeuchi, S., W. Zhang, K. Wakamatsu, S. Ito, V. J. Hearing, K. H. Kraemer and D. E. Brash (2004) Melanin acts as a potent UVB photosensitizer to cause an atypical mode of cell death in murine skin. Proc. Natl Acad. Sci. USA 101, 15076-15081.

75. Samokhvalov, A., L. Hong, Y. Liu, J. Garguilo, R. J. Nemanich, G. S. Edwards and J. D. Simon (2005) Oxidative potentials of human eumelanosomes and pheomelanosomes. Photochem. Photobiol. 81, 145148.

76. Ye, T., L. Hong, J. Garguilo, A. Pawlak, G. S. Edwards, R. J. Nemanich, T. Sarna and J. D. Simon (2006) Photoionization thresholds of melanins obtained free-electron laser photoelectron emission microscopy, femtosecond transient absorption spectroscopy, and EPR measurements of oxygen photoconsumption. Photochem. Photobiol. 82, 733-737.

Dan-Ning Hu*1,2, John D. Simon3 and Tadeusz Sarna4

1 Tissue Culture Center, Department of Pathology, The New York Eye and Ear Infirmary and New York Medical College, New York, NY

2 Department of Ophthalmology, Show Chwan Memorial Hospital, Taiwan

3 Department of Chemistry, Duke University, Durham, NC

4 Department of Biophysics, Jagiellonian University, Krakow, Poland

Received 29 October 2007, accepted 4 January 2008, DOI: 10.1111/ j.1751-1097.2008.00316.x

[dagger] This invited paper is part of the Symposium-in-Print: Melanins.

* Corresponding author email: (Dan-Ning Hu)

(c) 2008 The Authors. Journal Compilation. The American Society of Photobiology 0031-8655/08

Copyright American Society for Photobiology May/Jun 2008

(c) 2008 Photochemistry and Photobiology. Provided by ProQuest Information and Learning. All rights Reserved.

COMMENTS

0 comments

0 Comments



The information provided is no substitite for an informed medical professional. Please consult an expert before taking any action

POPULAR



June 5, 2019 Lip Service: How Contagious Are Cold Sores



June 4, 2019 Once Bitten, Twice Shy: How to Stop Mosquito Bites from Itching



June 3, 2019 Why Do We Sneeze? Your Email

SIGN UP

Advertising About Us Contact US Privacy Statement Terms of Service

Abuse Reporting Jobs

© 2002-2018 redOrbit.com. All rights reserved